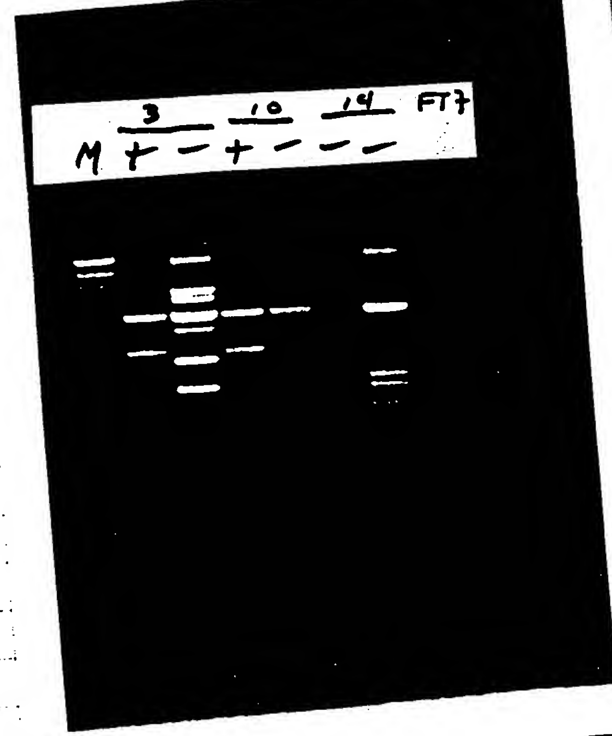
Exhibit 3

7/22 Digest FT7 cDNA 3, 10, 14 w/ KpnI/HindIII
use previous prep as control (-)



Expected size

	③ Sense	④ AntiS. ⊖
3	3159	3159
	1573	1366
	625	782
10	3159	3159
	1573	1121
	650	832
	380	650
14	3159	3159
	1573	945
	650	832
	204	650

Major problem w/ #14

Somehow samples got mixed-up

Go back & check out — Streak for single colonies
A B & E
Also mini prep from original culture
Digest w/ KpnI/HindIII O/N

7/23 Run gel of Digests
Clearly Sample A which was grown as maxi prep is in the wrong orientation
Start O/N of D & E to mini prep before starting 500ml culture

7/24 Mini prep FT7 cDNA 14 - D & E
Digest w/ KpnI/HindIII
run on gel - Both are fine
use D for maxi

7/25 Plasmid isolation of FT7, cDNA 14 - D up to Binding
- do 500ml in 2 250 sets, end w/ 2 tubes to Bind

7/26 Pull Bands - double band 1 prep - 6 hrs during the Day

7/27 Digest FT7-cDNA14 w/ KpNI/HindIII
① - Single band
② Double Binding
③ FT7-cDNA14 antisense

Check Absorbance / Concentration of

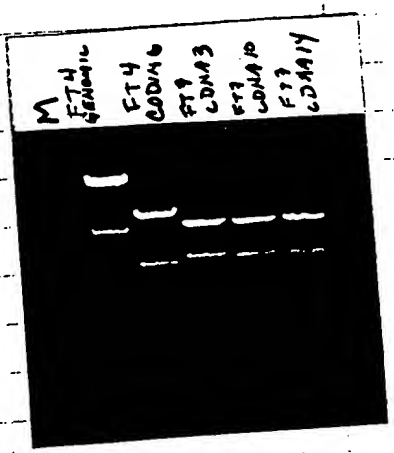
Samples	260 FT7 cDNA 14	280		260/280
①	.127	.062	.63 mg/μl	2.0
②	.143	.065	.71 mg/μl	2.3

Sequence
FT7, cDNA3
T7
cDNA-14
T7
8850
9007
8874

Digest Mouse FT4

	260	280	260/280
PssI genomic	.085	.045	
NheI/XbaI Coding	.106	.060	

Digest of cDNA3
KpnI/HindIII cDNA10
cDNA14



8/4 FACS Analysis of Transfected cells w/ following Vectors

pcDNA 7
FI7, 1,2a 2b, 3
1, 2b, 3
1, 3
cDNA 3
cDNA 10
cDNA 14

2 plates/ Vector — Divide
FACS 7 mls
Yuko 5 mls
CAT 3 mls (75 µl)
ETassy 5 mls (100 µl)

Antibodies

IgM	H	- black	1:100
IgM	Lx	green	1:1000
IgM	sLx		1:200
IgG	La	red	1:500
IgG	sLa	blue	1:500

2nd Antibody
IgM 2.5 mls
12.5/2.5 mls
IgG 1.5
6.0/1.5 mls

Results are
H - all neg
Lx - all neg
sLx, pcD (-), 1,2a 2b, 3(+), 1, 2b, 3(+), 1, 3 (-), cDNA 3 (-), 10 (+), 14(
La all neg
sLa all neg

8/8 Sequence 12 premier KG FT4

6451	6080
2470	6199
6374	6087
6306	6086
6203	6085
5721	5671

8/9 Run seq gel of above samples
Also sequence

5728	6084	7213
5731	5737	6082
5662	5727	6201
	5670	6200

8/10 Sequencing gel of 8/9 samples

Sequence

FT4 6079	cDNA 14
6202	T7
6307	8850
6373	8807
cDNA10 715	8874
946	

Protein assay of FACS Samples, Also CAT assay

	BSA	Blank	Protein	Ā
pcDNA	0	.100	.116	.108
FT7 1,2a2b,3	1	.204	.225	.214
FT7 1,2b,3	2	.369	.383	.376
FT7 1,3	4	.691	.673	.682
cDNA3	8	1.215	1.230	1.222
cDNA10	16			
cDNA14				

Sol'n/Sample			Ā
pcDNA	.292	.294	.293
1,2a2b,3	.337	.330	.33
1,2b,3	.298	.343	.32
1,3	.369	.372	.37
cDNA3	.363	.379	.3
cDNA10	.306	.298	.3
cDNA14	.225	.253	.2

FACS Results

Only stain w/ sh x

1,2a2b,3	23.6%
1,2b,3	24.6%
cDNA10	14.9%
cDNA14	8.0%

8/11 Sequencing of FT4 9/10 Samples

Micro BCA Protein Assay [redacted]

Reagent mix	MC	MB	MA
Per assay tube (ml)	0.01	0.24	0.25
Cocktail for Tubes			

Incubate 1 h at 60¡C and cool to room temp.
Since the color development has no end point, all tubes must be heated and cooled at the same time

1 mg/ml BSA (l)	Water (l)	Reagent (l)	Abs. 562	
0.0	500.0	500.0	Blank	Slope = 0.0734
1.0	499.0	500.0	0.108	Y intercept = 0.0656
2.0	498.0	500.0	0.214	X intercept = -0.8940
4.0	496.0	500.0	0.376	R = 0.9985
8.0	492.0	500.0	0.682	
16.0	484.0	500.0	1.222	

Sample	l in assay	Water (l)	Reagent (l)	Abs. 562	mg protein/ml
pcDNAI	5.00	495.00	500	0.293	0.62
FT7 1,2a2b,3	5.00	495.00	500	0.333	0.73
FT7 1,2b,3	5.00	495.00	500	0.320	0.69
FT7 1,3	5.00	495.00	500	0.370	0.83
cDNA 3	5.00	495.00	500	0.371	0.83
cDNA 10	5.00	495.00	500	0.302	0.64
cDNA 14	5.00	495.00	500	0.239	0.47

Cocktail 15
3H Chloe 300
Tris, 2M pH 8.0 75
But CoA 75
H_2O 300
50/tube

8/11 CAT Assay (FT7) samples
2.5 μl of cell extract
Control
pcDNA
1,2a2b,3
1,2b,3
1,3
cDNA 3
cDNA 10
cDNA 14

CAT [redacted]

Vector		Counts/5ul 8/12/94		Incorporated Counts (.95)		Total Counts		Total Counts Incorporated	
pcDNAI		11,349	11,829	9,169	8,063	236,169	244,643	9,673	8,487
FT7 1,2a2b,3		11,161	11,441	27,211	21,919	250,431	250,739	28,643	23,073
FT7 1,2b,3		11,772	11,826	37,541	40,684	272,981	277,204	39,517	42,825
FT7 1,3		11,215	11,690	23,076	28,706	247,376	262,506	24,291	30,217
cDNA 3		11,834	11,206	33,885	39,096	270,565	263,216	35,668	41,154
cDNA 10		12,017	11,312	30,066	33,165	270,406	259,405	31,648	34,911
cDNA14		11,079	11,570	44,133	40,529	265,713	271,929	46,456	42,662
Control			10,354		424		207,504		446
	Protein Conc. (ug/2.5ul)	Total Counts Inc- Bkg		% INC/hr		% INC/hr/ug		Mean CAT Activity	
pcDNAI	1.55	9,249	8,063	3.92	3.30	2.53	2.13	2.33	
FT7 1,2a2b,3	1.83	28,219	22,649	11.27	9.03	6.17	4.95	5.56	
FT7 1,2b,3	1.72	39,093	42,401	14.32	15.30	8.33	8.89	8.61	
FT7 1,3	2.07	23,867	29,793	9.65	11.35	4.66	5.48	5.07	
cDNA 3	2.07	35,244	40,730	13.03	15.47	6.29	7.48	6.88	
cDNA 10	1.60	31,224	34,487	11.55	13.29	7.22	8.31	7.76	
cDNA14	1.17	46,032	42,238	17.32	15.53	14.81	13.28	14.04	

8/12 Assemble Data of FT7 FACS, CAT Assay / give to Jacko
Work on FT4 sequence

8/15 7 deaza sequencing on Troublesome FT4 samples

6451	6200
6378	6079
6306	6307
6203	1897
5721	1898
	1899

8/16 Sequencing gel of 8/15 samples (FT4) Formamide gel
Probe Thomas's northern Blot w/ GAP probe
To check condition of RNA

8/18 The 7 deaza technique didn't resolve all of the compressions
Try a Terminal transferase technique
Run standard sequenase rxn, after extension reaction
Heat tubes (A, C, G, T) for 1.5 mins 100°C
Hold on ice 10 min, Prepare TdT/dNTP cocktail
Add to tubes, 37°C 30 min
Add 5 μl Stop